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Advancements in spectroscopic and microscopic techniques for modern analysis

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ABSTRACT

Nowadays, different well-established analytical techniques are employed for qualitative and quantitative analysis of a wide variety of samples. The use of instrumentation by chemists is ubiquitous in research. The material properties such as crystal structure, surface state, surface morphology, composition, optical and magnetic characteristics probed is an important step in understanding their chemical and physical performance. These instruments provide information about the surface characterization and structural elucidation of a sample. A detailed description of each instrumentation technique is usually linked with its associated theory, design and application.

The study entailed the important factors to consider when selecting suitable instrumentation for analysis, whether destructive or non-destructive and highlighted merits and demerits. The techniques such as Fourier Transformed Infrared (FT-IR) Spectroscopy, Scanning Electron Microscope-Energy Dispersive X-Ray (SEM-EDX), Atomic Absorption Spectroscopy (AAS), X-Ray Diffraction (XRD), UV-Visible Spectroscopy, Nuclear Magnetic Resonance (NMR) Spectroscopy, Inductively Coupled Plasma-Optical Emission Spectroscopy (ICP-OES), Transmission Electron Microscopy (TEM) and X-Ray Fluorescence (XRF) Spectroscopy are reviewed. These insights will inform the researchers with a useful guide to assist them on the choice the appropriate instrumentation technique for their analytical research.

Keywords: analysis, elucidation, spectroscopy, instruments, characterization, microscopy

INTRODUCTION

Modern analytical chemistry is a dynamic field that deals with the principles and concepts of important spectroscopic, microscopic and spectrophotometric techniques for surface and structural analysis (Bisen and Sharma, 2013; Wang *et al.*, 2015). The techniques are pivotal in diverse fields such as medicine, food science, geology, nanotechnologies, materials science, and environmental analysis (Vickerman and Gilmore, 2009; Robinson *et al.*, 2014). Therefore, development, intercomparisons and applications of instrumentation techniques are valuable for analysis in scientific research, control or industrial laboratories.

A description of each technique is associated with its related theory, instrumentation design, and application in various fields (Rayson, 2004; Warner *et al.*, 2016). The instruments are usually interfaced with a computer, as a readout device with some instruments fully automated to perform all the functions (Kovarik *et al.*, 2020; Shah *et al.*, 2020). These instruments are highly selective and sensitive and are therefore important in different fields (Farmer and Jimenez, 2010). The nature of materials such as the composition, surface state, morphology, crystal structure, optical and magnetic characteristics probed is important in understanding their physical and chemical performance (Liu *et al.*, 2014). The instruments used can be coupled with other instruments to provide more vital information about the surface characterization and structural elucidation of the sample of interest (Kumar *et al.*, 2014). However, the main challenges for modern analysis that needs to be addressed is the pricy cost of the instruments, their complexity and accessibility.

The instruments such as Fourier Transformed Infrared (FT-IR) Spectroscopy (Tinega *et al.*, 2023), Scanning Electron Microscope-Energy Dispersive X-Ray (SEM-EDX) (Hummadi *et al.*, 2022; Rosanti *et al.*, 2022), Atomic Force Microscopy (AFM) (Deng *et al.*, 2018), X-Ray Diffraction (XRD) (Njeri *et al.*, 2023), UV-Visible Spectroscopy (Oke and Mohan, 2022), Nuclear Magnetic Resonance (NMR) Spectroscopy (Jaime *et al.*, 2018), Atomic Absorption Spectroscopy (AAS) (Ndung'u *et al.*, 2020), Transmission Electron Microscopy (TEM) (Karthikeyan *et al.*, 2016), Inductively Coupled Plasma-Optical Emission Spectroscopy (ICP-OES) (Khan *et al.*, 2015) and X-Ray Fluorescence (XRF) Spectroscopy (Ndung'u *et al.*, 2023) among others are used for analysis. However, reports indicate limited information on their working principle, emerging innovations and applications of techniques and this formed the basis of this review work.

SPECTROSCOPIC TECHNIQUES

The techniques are employed to measure matter interaction with electromagnetic radiations of different frequencies (Akash and Rehman, 2020). The magnitude of the energies involved, magnetic field and instrumentation considerations has led to the rise of different spectroscopic techniques (Bisen and Sharma, 2013). The FT-IR, AAS, XRD, UV-Visible, XRF and NMR techniques are reviewed in the following sub-sections.

Atomic Absorption Spectroscopy (AAS)

The technique uses absorption spectrometry to examine the analyte concentration in a sample (Potts *et al.*, 2004; Ivanenko *et al.*, 2012). It entails absorption of

electromagnetic energy of a certain wavelength, mainly ultraviolet or visible region, by ground atoms in their gaseous state (Farrukh, 2012). It involves absorption of light by free atoms of an element at a wavelength specific to that element therefore capable of quantifying sample chemical entities in trace elemental analysis (M Series, 2005; Wilberforce, 2016). The technique is very sensitive as its detection limit can be as low as 1 ppm using the flame procedure, with spectral lines extremely narrow (0.02-0.5Å) and transition energies unique for each element (Paul *et al.*, 2014, Paul *et al.*, 2017). The technique is employed in the determination of metals in vast samples like biomaterials, food, nanomaterials, drugs, environmental, forensics and industrial wastes (Farrukh, 2012).

Generally, AAS instrument consist of a stable continuous radiation source, lenses, mirror and a slit, monochromator for selecting narrow wavelength band from the source; a sample compartment for holding the sample being analyzed; a detector and a readout device (Sudunagunta *et al.* 2012; Wilberforce, 2016) as shown in Figure 1. The sample is converted into atomic vapour and then absorption specific for a given element is measured (Paul *et al.*, 2017). The sample element is aspirated to a flame, then changed into atomic vapour (Yeung *et al.*, 2017). Some atoms are thermally excited by the flame whereas most of them remain in the ground state (Wilberforce, 2016). The ground state atoms absorb a specific wavelength radiation (Akash and Rehman, 2020). The amount absorbed corresponds to metal ions concentration (Sharma and Tyagi, 2013). It's worth noting that the hollow cathode lamp used is specific to the metal being analyzed.

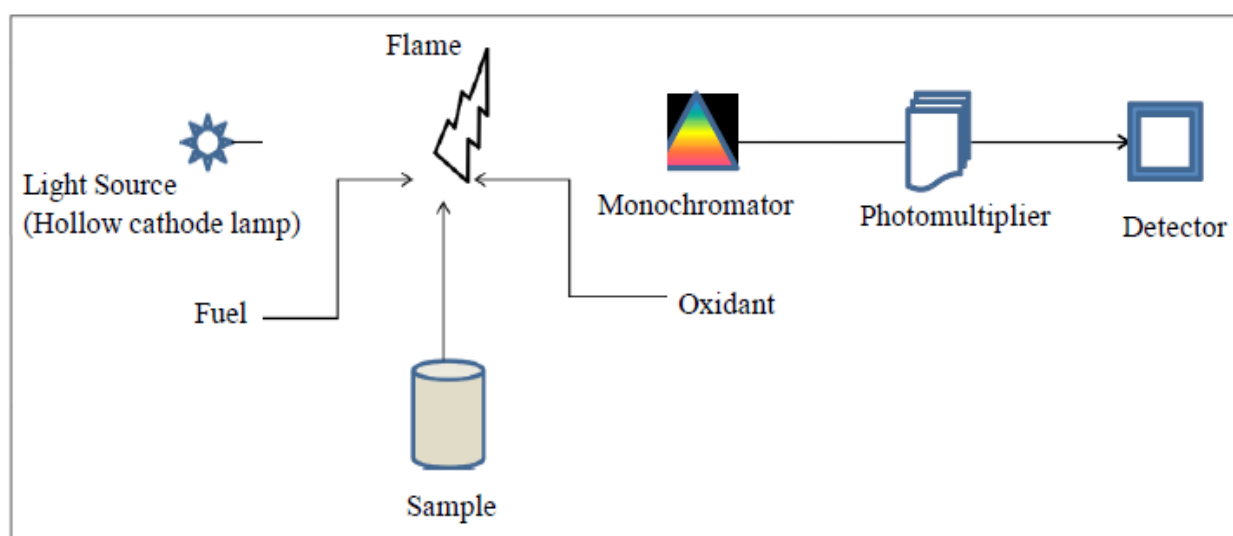


Figure 1: Components of Atomic Absorption Spectrophotometer (AAS)

Source: (Paul *et al.*, 2017)

Fourier Transform Infra-Red (FT-IR) Spectroscopy

The spectroscopic analysis in the mid-infrared spectral region (4000 cm^{-1} - 400 cm^{-1}) provides quantitative structural, chemical, and compositional information on constituent molecules in the solid, gas and liquid phases, thus employed in vast field of industrial applications such as in medicine, materials science, biotechnology and environmental analysis (Haas and Mizaikoff, 2016). It is a non-destructive, rapid and time saving technique that is able to detect a range of functional groups and is sensitive to changes in molecular structures (Amir *et al.*, 2013; Kurniawati *et al.*, 2021; Rosanti *et al.*, 2022). FT-IR provides vital information on the chemical constitution and physical state of a sample (Cocchi *et al.*, 2004). The techniques determine the chemical bonding, molecular structure mainly functional groups of organic and inorganic materials (Munajad *et al.*, 2018; Al-Ghouti and Al-Absi, 2020). The technique has merits over other analytical techniques such as a small sample size use (milligram range), easy sample preparation, accurate, fast analysis, inexpensive and reliable and a simple sampling procedure (Sivakumar *et al.*, 2012; Chen *et al.*, 2014). It is widely used in many laboratories due to its improved

signal-to-noise ratio and fast operational speed (Cui *et al.*, 2017).

The technique characterizes functional groups bond structure after IR radiation absorption causing dipole moment a change (Țucureanu *et al.*, 2016). This causes molecular bond vibrations and rotations resulting to a FT-IR spectrum at varying wavenumbers in the IR region (Faghihzadeh *et al.*, 2016; Perez-Guaita *et al.*, 2016). These FT-IR spectra exhibit characteristic absorption frequencies of varied intensities for different functional groups in a sample (Santhoshkumar *et al.*, 2017; Gonciarz *et al.*, 2020). In the modern analysis, advancements have resulted to rise of more digitalized FT-IR instruments that are faster and more sensitive as they can detect vast compounds (Simonescu, 2012).

The working principle of FT-IR (Figure 2) is that the light emitted is split to two beams (Valand *et al.*, 2019). Then, the light beams are re-combined again using fixed and movable mirrors (Cui *et al.*, 2017). The combined beams interact with the sample under study and the detector measures the radiation intensity moving into a sample and the radiation intensity transmitting through the sample (Munajad *et al.*, 2018).

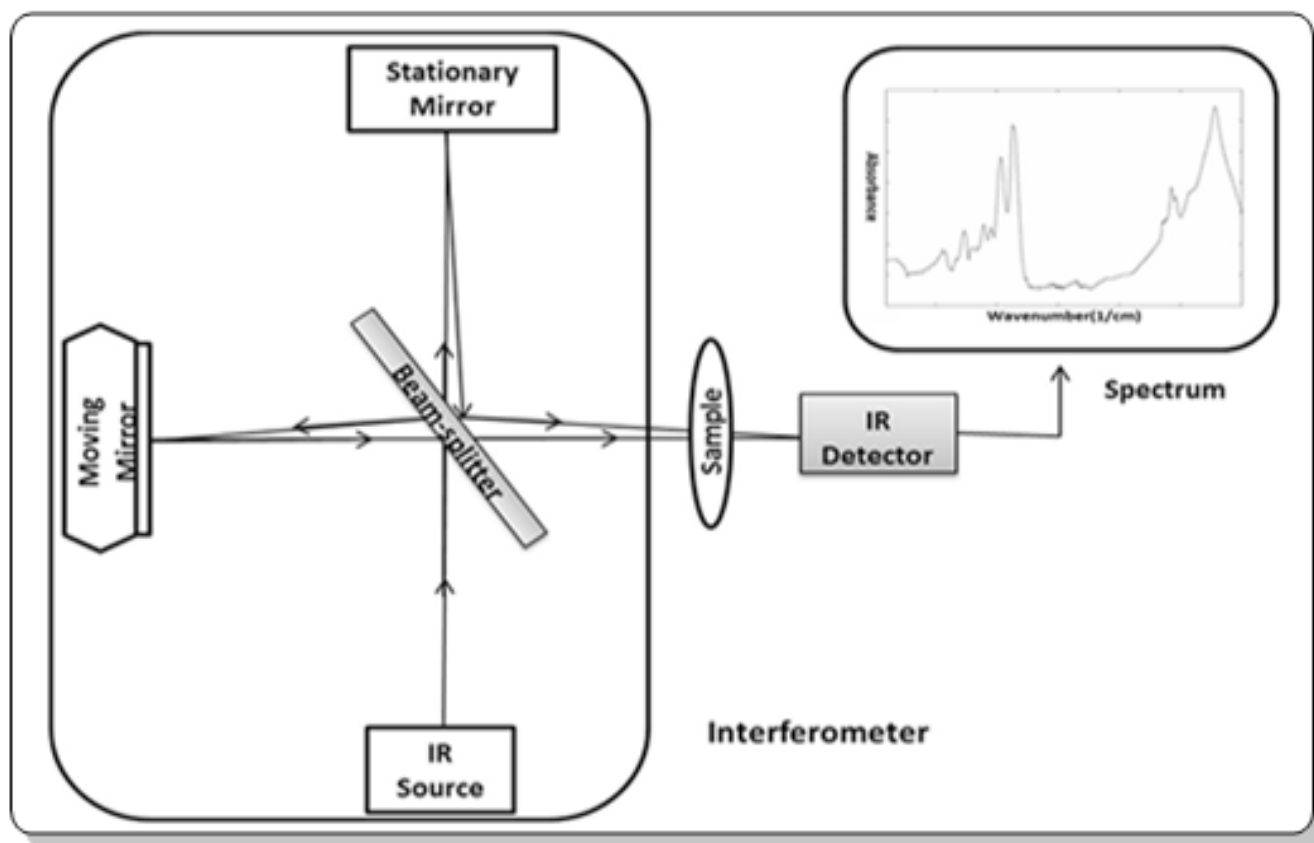


Figure 2: The FT-IR instrument components

Source: Faghihzadeh *et al.* (2016)

UV-Visible (UV-Vis) Spectroscopy

The technique measures the light intensity in the ultra-violet (10–400 nm) and the visible (400–800 nm) regions as a function of wavelength expressed in nanometers (Akash and Rehman, 2020). It is the most employed technique as an inexpensive, rapid, simple, flexible, simple, accurate, precise, non-destructive, specific and applicable to small quantities of compounds (Chakraborty *et al.*, 2018; Singh, 2020). The ultra-violet (UV) spectroscopy utilizes light in the ultraviolet, visible and Infra-Red ranges (Picollo *et al.*, 2018; Verma and Mishra, 2018). The technique measures the absorbance of light that passes through the test sample as a function of wavelength dependent on the molecules present (Singh, 2020).

This technique is centered on transition of valence electrons from their ground state to an excited state driven by electromagnetic radiation in the UV–Vis range (Haddi *et al.*, 2024). The light absorbed provides information about electronic transitions that occur within the sample

material (Guo *et al.*, 2020). The radiation sources are deuterium or hydrogen discharge tube with 185 nm to 390 nm wavelength range, and tungsten-filament lamp which covers a visible region of 350 nm to 800 nm (Abu Bakar *et al.*, 2016). The UV-Vis spectrophotometers measures the light absorbed or transmitted as a function of wavelength (Rocha *et al.*, 2018).

The working principle of the technique (Figure 3) is based on passing a monochromatic radiation through the sample solution where the emitted radiation intensity depends on the thickness and the sample concentration (Shinde *et al.*, 2020). This is as explained by the Beer-Lambert law, which explains that the interaction between light that interacts with the sample (I , transmittance or reflectance) versus the incident intensity (I_0) depends on the concentration of the absorbing species in the solution and the path length (Verma and Mishra, 2018). Typically, the results are graphically presented as absorbance (no units) against wavelength in nanometers (nm) (Rocha *et al.*, 2018).

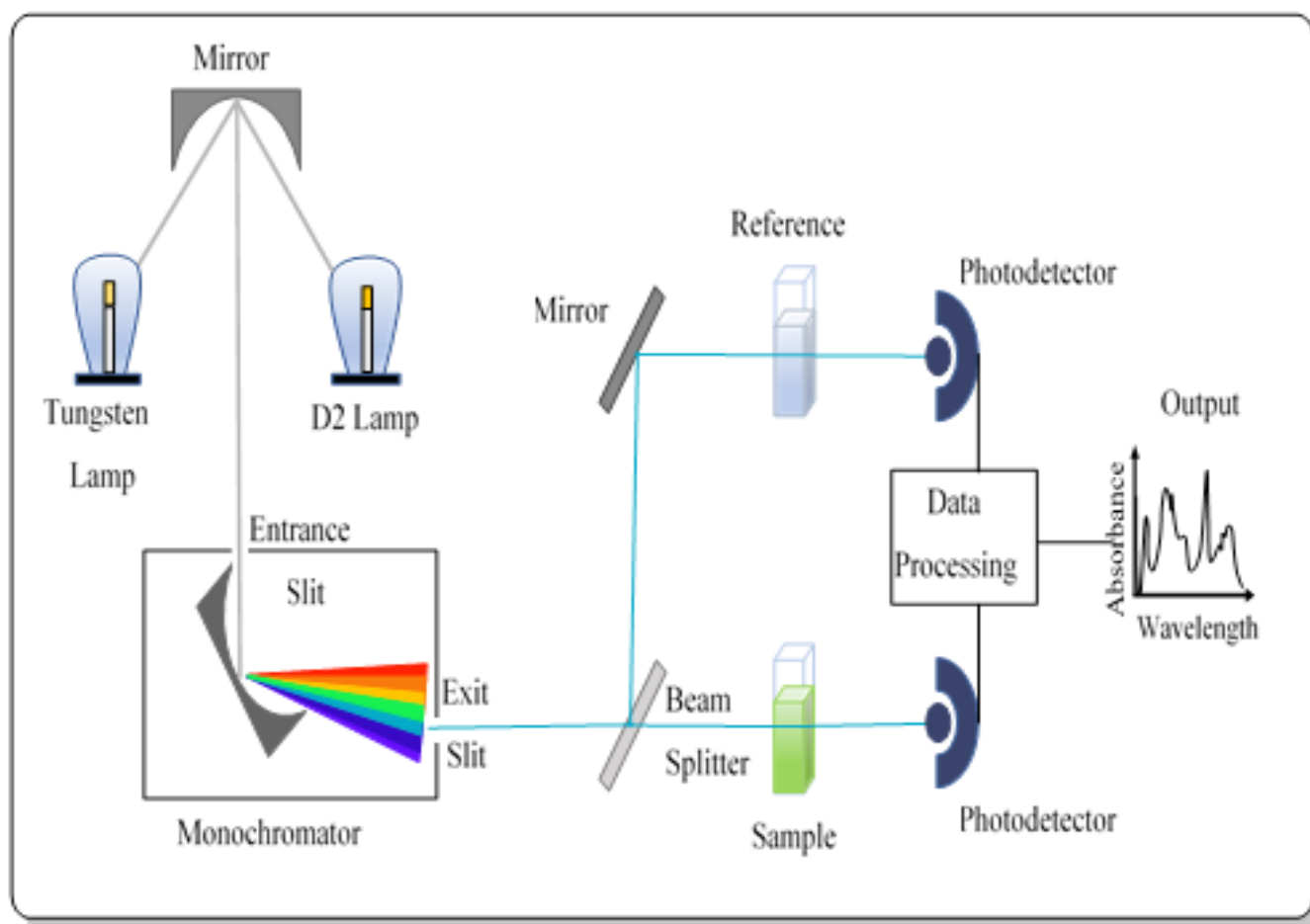


Figure 3: Components of UV-Visible spectrophotometer

Source: Rocha *et al.* (2018)

X-Ray Diffraction Spectroscopy

The spectroscopy is a non-destructive and a simple technique that offers a quick quantitative and qualitative

analysis of pure and multi-component compounds that requires minimal sample preparation (Khan *et al.*, 2020). It is used in determining the elemental proportions of

the sample under study and its structural state (Bishnoi *et al.*, 2017; Titus *et al.*, 2019; Fakhirah *et al.*, 2024). The technique determines the crystalline structure, phase nature and crystallite grain size of sample materials (Melkamu and Feleke, 2022; El-Azazy *et al.*, 2023; Pandey *et al.*, 2021; Ermrich and Oppen, 2011).

The principle is founded on constructive interferences of the monochromatic X-rays with the sample (Bunaciu *et al.*, 2015). The X-rays from the cathode ray tube are filtered producing monochromatic radiations which are then collimated and directed to the sample being analyzed

(Figure 4) (Epp, 2016). The positions of the diffracted incident X-Ray beams are ruled by atoms positions in the crystal unit cell (Kim *et al.*, 2013). This phenomenon is explained by Bragg's law which describes the relationship between the incident X-Ray light and its reflection from the crystal surface (Figure 5) (Cornelius and Thomas, 2018). The X-ray beam scatters off a planes at a certain angle, θ , which equals the incident angle with respect to crystalline planes (George and José, 2021). The diffracted X-rays are detected, processed, analyzed and counted (Bunaciu *et al.*, 2015).

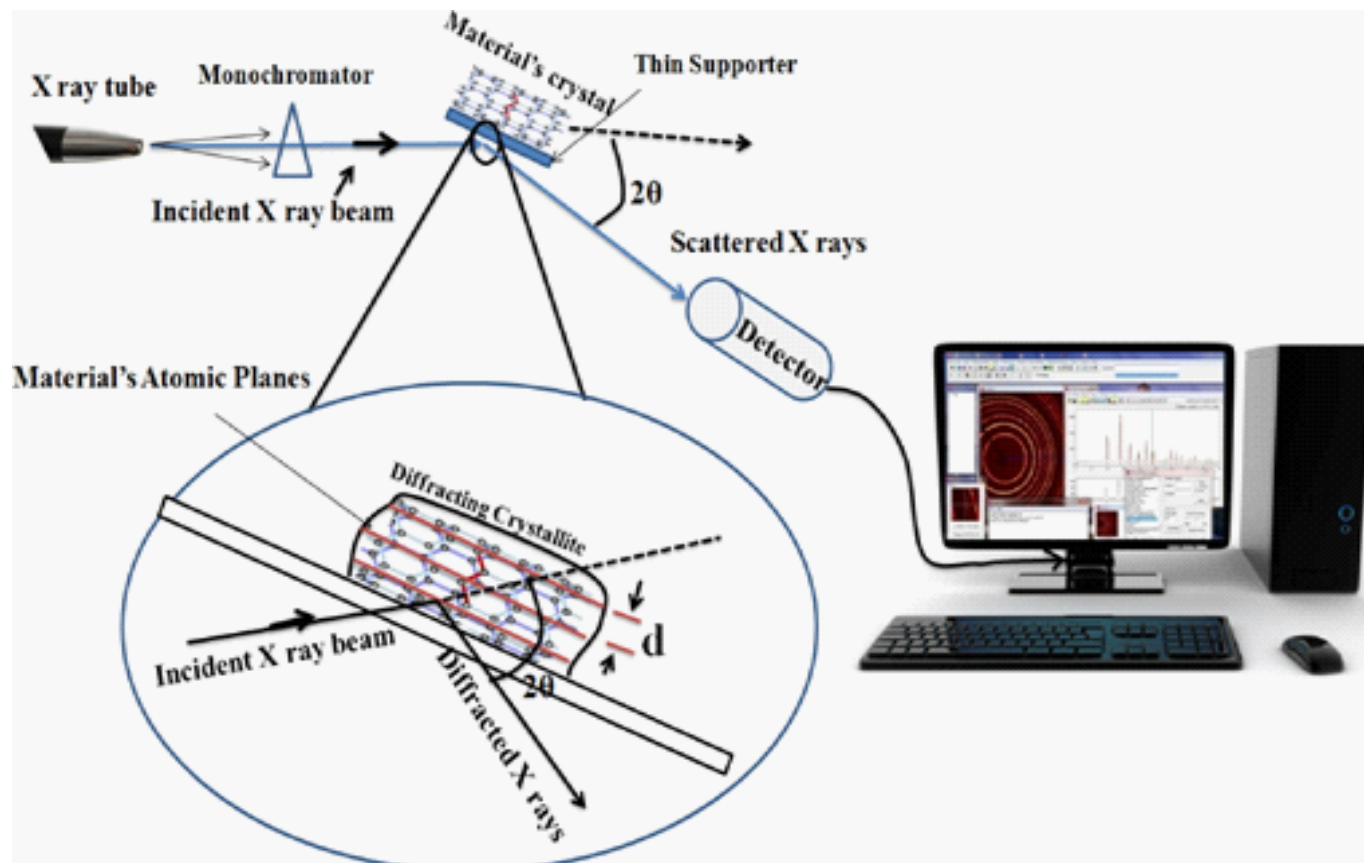


Figure 4: Components of an XRD instrument

Source: Das *et al.* (2014)

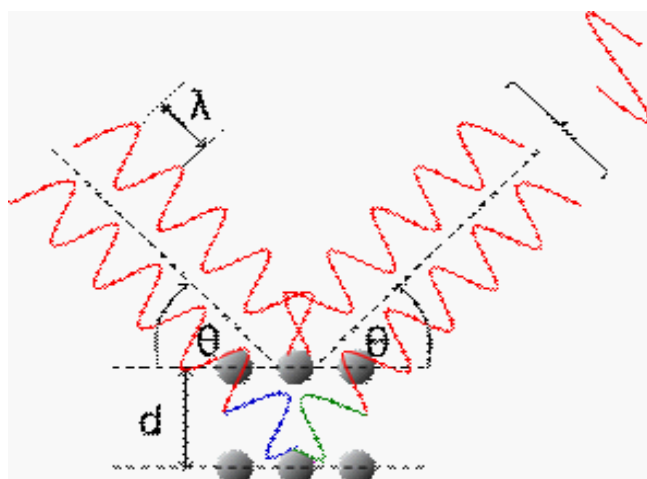


Figure 5: Bragg's law diagram

Source: Plaipichit *et al.* (2018)

The crystallite size of crystalline samples is usually determined using Scherrer equation (Muniz *et al.*, 2016; Chen *et al.*, 2023) expressed by Equation 1.

$$L = \frac{k\lambda}{\beta \cos \theta} \quad (1)$$

Where L is the average crystal size (nm), k is the shape factor constant (0.9), λ is the specific wavelength of the X-ray used (1.5406), θ is the diffraction angle and β is the angular width in radius.

X-Ray Fluorescence (XRF) Spectroscopy

The analytical tool is employed for qualitative and quantitative analysis of multi-elemental components

(Grieken and Margui', 2013). It is a versatile tool in trace elements analysis (Potter and Brand, 2019). The technique is based on the wavelength-dispersive principle, which entails emission of X-Ray photon energy of individual atoms that can be measured (Oyedotun, 2018). It is an established technique in application in many fields, such as in the biological, archeological, environmental and forensic sciences as well as in industry (Tsuji *et al.*, 2015; Wilberforce, 2016).

The principle is based on the use of an X-ray beam to produce ionisation in the atoms inner shells present in the sample due to photoelectric absorption (Marguí

and Grieken, 2013). It is centered on the effect of X-ray interactions with the matter, causing excitation of certain electrons, which when relax to their ground state, emitting X-rays of a wavelength characteristic to a particular element (Gupta, 2014). The energy calibrations are first carried out to ensure that peak energies were accurately tied to specific elements. The XRF analysis is done by measuring the energy of fluorescent photons emitted by a sample after being excited on X-ray beam (Montanha *et al.*, 2020). The emitted X-Ray energy or wavelength is then employed to identify the different elements present in the sample which is proportional to its concentration (Marguí *et al.*, 2014). The Figure 6 is a schematic diagram of XRF spectrometer.

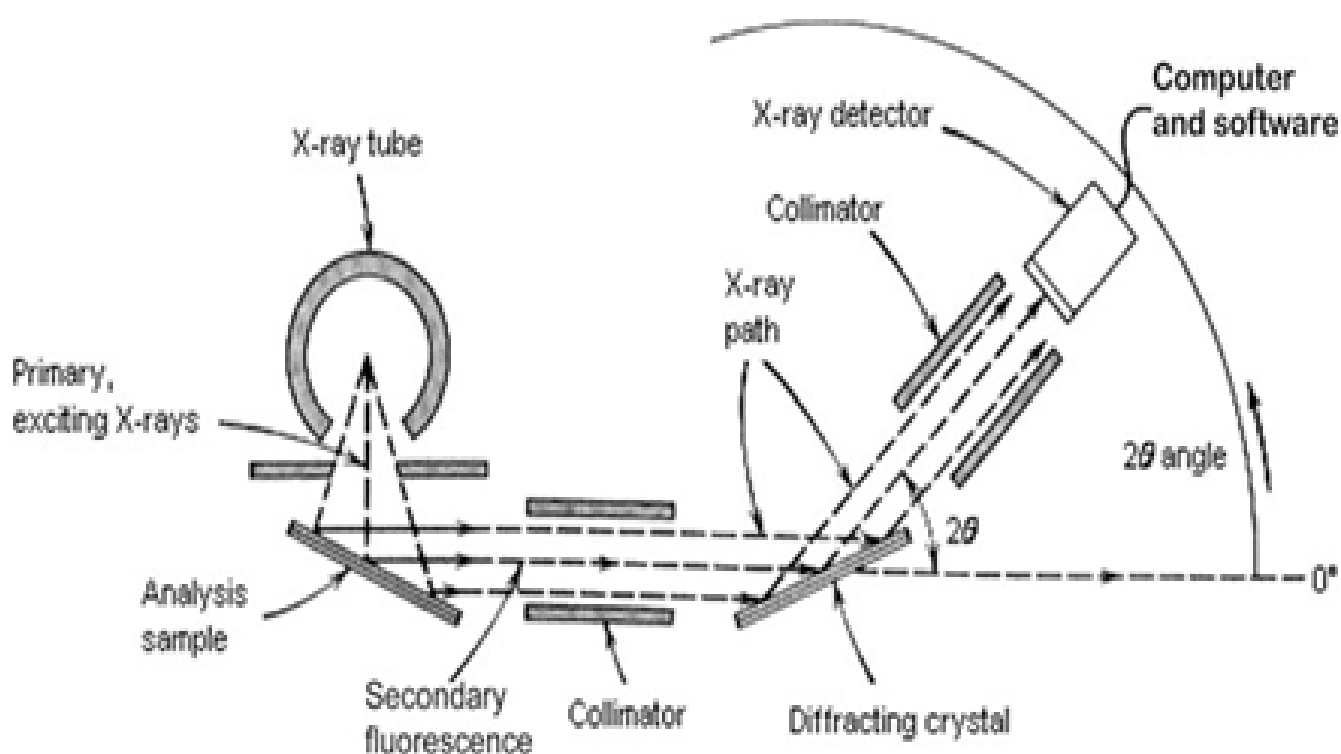


Figure 6: X-Ray Fluorescence spectrometer

Source: Shackley (2011)

Nuclear Magnetic Resonance (NMR) Spectroscopy

This is the most powerful technique in the modern science that utilizes radiofrequency waves to reveal information about magnetic nuclei of a compound (Bothwell and Griffin, 2011). It is a simple and a non-destructive technique that requires a small sample size and provides structural information that is quantitative (Claridge, 2016). The NMR technique has a broad scope in application such as in pharmaceuticals, organic chemistry, food science, biochemistry and material science (Yu *et al.*, 2021). It is an informative tool that is employed to determine the chemical makeup of organic molecules

(Muller *et al.*, 2014). It is employed in vast fields due to its versatility in structural elucidation and the interactions in a range of molecular systems (Simpson *et al.*, 2018).

The technique utilizes permanent magnets that generate magnetic fields and therefore offering advantages of simplicity in operation and cost-effective, and this offers a viable alternative to high-field NMR spectroscopy (Yu *et al.*, 2021). The frequencies of NMR transitions are in the range of 10-1000 MHz which probe the nuclei of atoms. The nuclei show an energy state change when the sample analyte is placed in an intense magnetic field (Mishra *et*

al., 2017). The NMR signal produced reflect the chemical environment and the dynamics of a nuclear spin (Sugiki *et al.*, 2017). This is referred to as the “chemical shift” and it identifies different chemical groups within a molecule and their chemical bond environment (Simpson *et al.*, 2012). The NMR analyst can easily determine the degree of purity in a sample being analyzed, and this practice is used to validate the purity of different synthetic standards (Espina *et al.*, 2009).

The basic principle is that the chemical and structural constitution of a sample is determined by their nuclei which have distinctive magnetic fields (Kong *et al.*, 2015). The NMR spectrometer performs analysis using magnetic fields and a special detector is utilized to assess the changes (Figure 7) (Zia *et al.*, 2019). The external magnetic field strength makes the electrically charged nucleus to transition from a lower to a higher energy level with the difference depended on the magnetic field power and the nuclear field moment (Keeler, 2011).

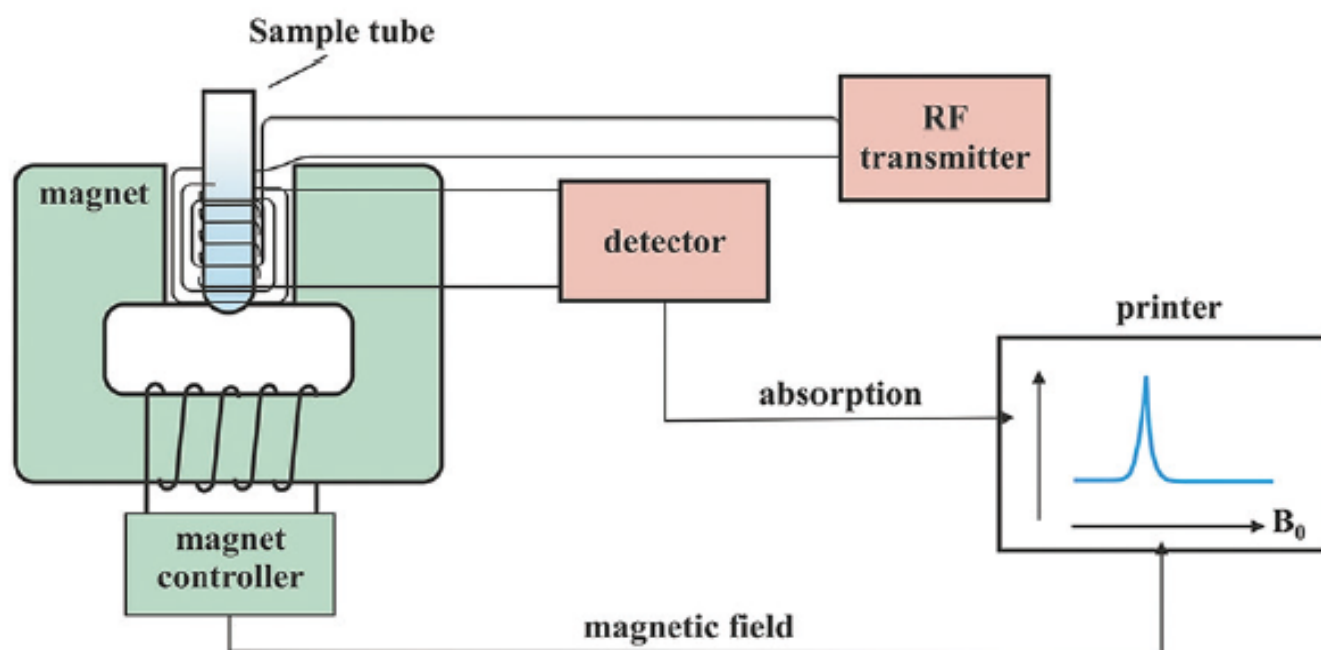


Figure 7: A schematic diagram of Nuclear Magnetic Resonance (NMR) spectrometer

MICROSCOPIC TECHNIQUES

Microscopy entails a “direct” imaging of the sample surface in visualizing the macro and/or microscopic structure and therefore provides the best resolution than the light-based imaging techniques (Kaeche, 2013). The electron microscopy is on the basis of a circular electromagnetic field which acts on electron beam in a way related to the action of a glass lens on a photons beam (Bisen and Sharma, 2013). The SEM-EDX and TEM techniques are reviewed in the following sub-sections.

Transmission Electron Microscopy (TEM)

The technique provides structural information on the materials (Franken *et al.*, 2017). The TEM image provides a detailed information about the nano-structural characteristics of a sample material (Geetanjali *et al.*,

2024). The technique depends on the electron beam energy (eV), microscope resolving power (smaller than 0.3 nm), specimen thickness (less than 1 μm), stability and composition (Goodhew, 2011).

The principle is that matter interacts with an electron beam producing X-rays which are then used for elemental analysis using specialized detectors (Kaeche, 2013). The transmitted electrons are focused by lenses and collected by parallel detectors to form an image (Inkson, 2016). The electron gun in a TEM instrument accelerates electrons of upto 300 kV so as to provide sufficient energy for them to penetrate through the sample material of up to 1 μm (Reimer and Kohl, 2008). The TEM microscope comprise of an electron gun, the electrostatic lenses which focusses electrons and a transmitted electron detection system (Figure 8).

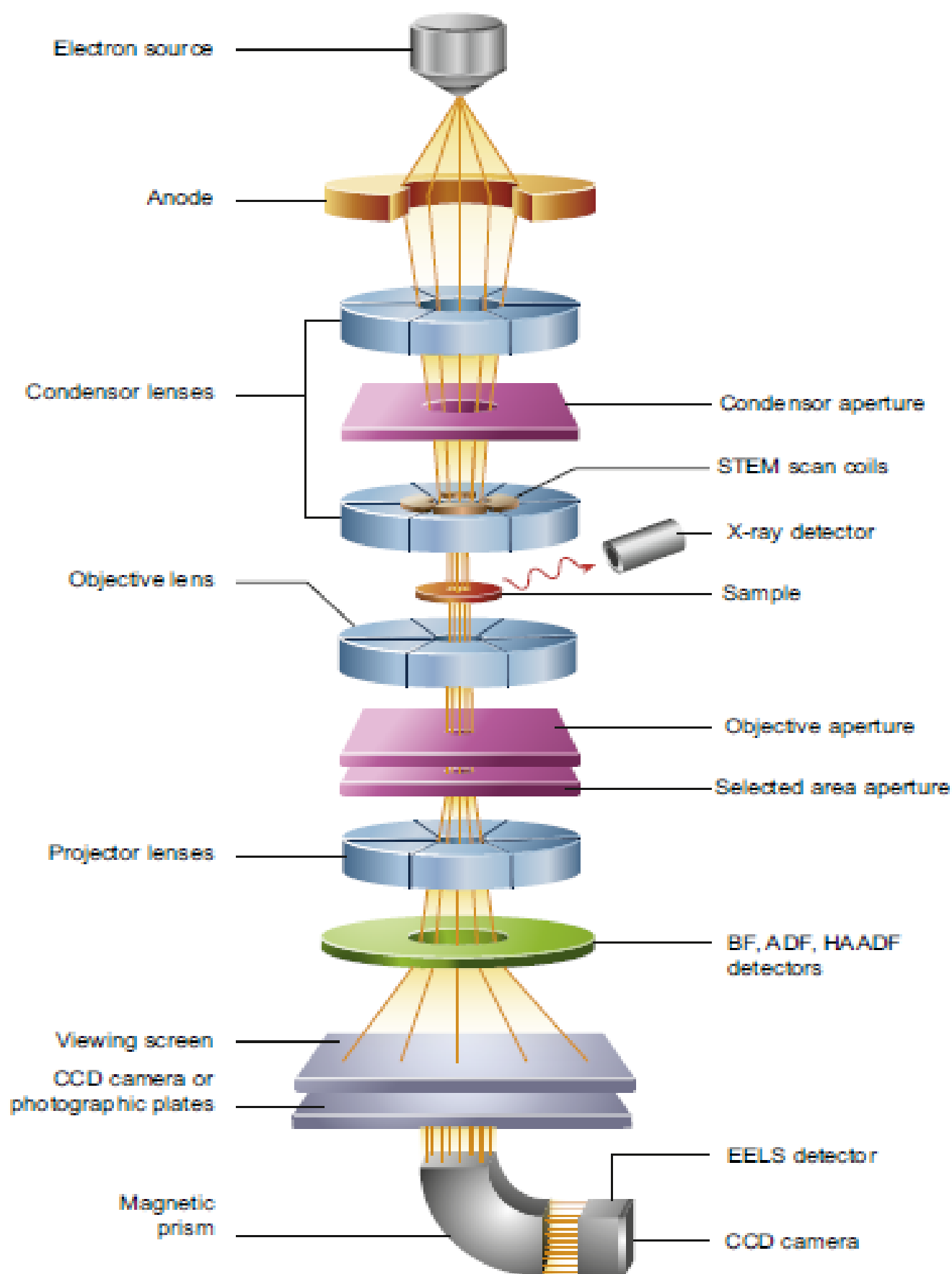


Figure 8: Schematic diagram of TEM components
Source: Inkson (2016)

Scanning Electron Microscope-Energy Dispersive X-Ray (SEM-EDX)

The microscopy uses beam of electrons which interacts with the sample and scatters to produce a signal which provides information about topological image and relative composition (Nie *et al.*, 2015; Han *et al.*, 2018). SEM images of various magnifications are employed to evaluate the morphology and sample size (Sharma *et al.*, 2016). The SEM instrument is mostly coupled together with Energy Dispersive X-Ray (EDX) detectors as an additional tool for semi-quantitative analysis (Cardell and Guerra, 2015; El-Zahhar *et al.*, 2019).

The working principle is that SEM uses a focused electron beam from an energy cathode which upon interacting with a sample produces a topological image and its relative chemical constitution (Haddi *et al.*, 2024). The focused electron beam produces secondary electrons, backscattered electrons and characteristic X-rays upon interaction with the sample material, which then generate SEM signals that are collected by proper detectors to form images (Abd Mutalib *et al.*, 2017; Han *et al.*, 2018). The SEM instrument constitutes an electron source, column containing electromagnetic lenses, sample chamber, electron detector and the computer display (Figure 9).

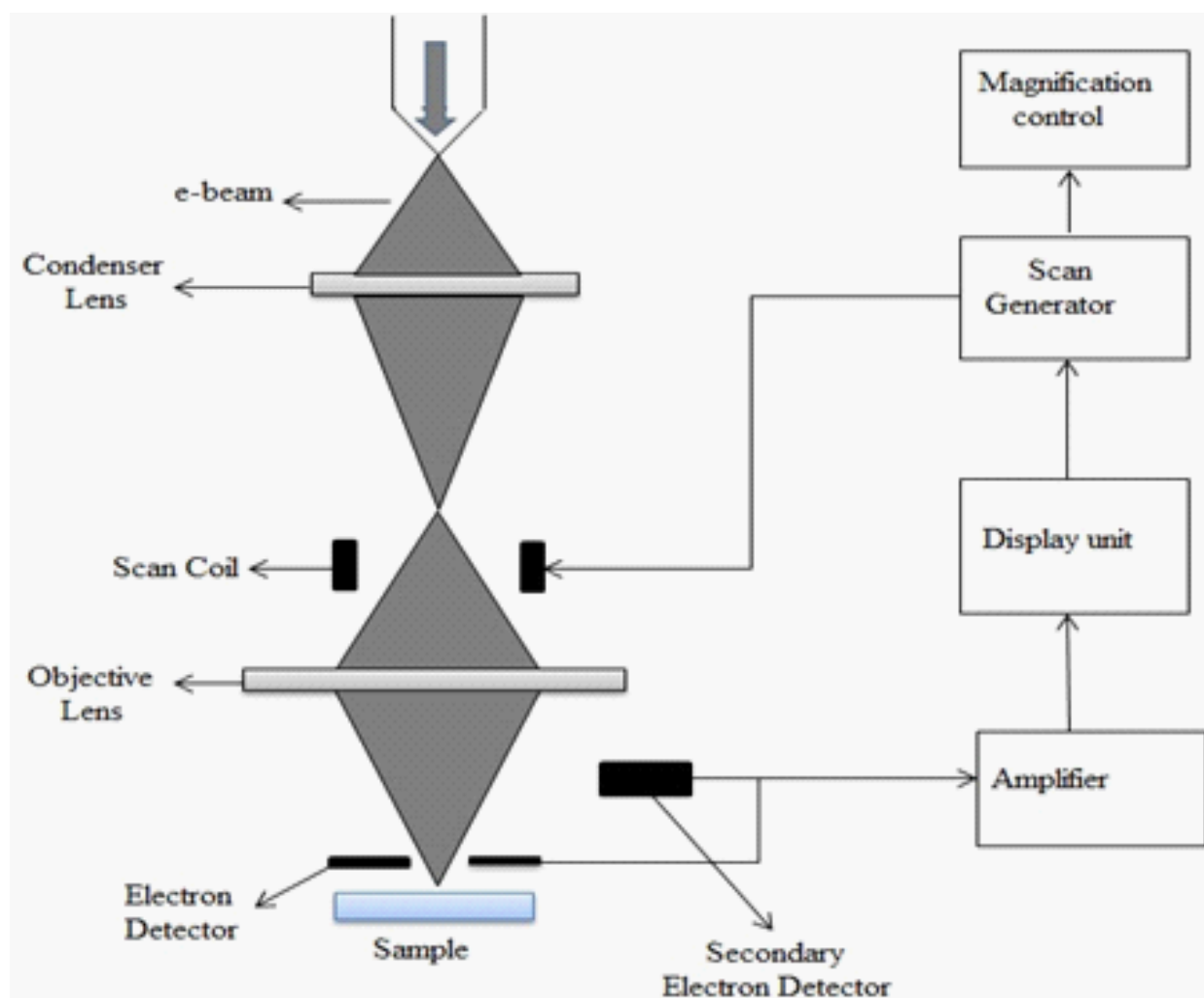


Figure 9: Schematic view of SEM components

Source: Han *et al.* (2018)

CONCLUSIONS

Spectroscopic and microscopic techniques are reported to be versatile and highly informative analytical tools for characterization and structural elucidation. The study reviewed FT-IR, AAS, XRD, UV-Visible, XRF and NMR techniques because of their increased usability by scientists in different fields such as in geology, medicine, food science, materials science, nanotechnologies, and environmental analysis amongst others. The techniques

provide vital information on the chemical composition of organic and inorganic moieties making them unique and ideal in analysis. The existence of more diverse techniques that complement spectroscopic and microscopic studies further attract a more advanced review to enhance their potential impact to analysts. The Table 1 shows a summary of the reviewed spectroscopic and microscopic techniques highlighting their merits, demerits and some of the applications.

Table 1: Summary of reviewed spectroscopic and microscopic techniques

S/No	Technique	Merits	Limitations	Applications
1	AAS	Low operational costs, simple and fast working, highly selective and sensitive, performs quantitative analysis of metals in trace amounts.	Cannot detect non-metals, sample is destroyed, spectral interference, cannot analyze metal oxides, not suitable for complex matrices	Environmental analysis, pharmaceuticals, geological analysis, food and beverage analysis, biochemical analysis
2	FT-IR	Rapid and speedy, accurate, non-destructive, high sensitive, small sample required, high resolution, rapid reproducibility of data, ease of use, spectral quality, sensitive	Limited surface sensitivity, expensive equipment in purchase and maintenance, requires careful sample preparation, water interference, limited identification	Food analysis, medical analysis, environmental analysis, drug analysis, mineral analysis
3	UV-Visible	Cost-effective, non-destructive, quick measurements, easy to use, versatile, can perform quantitative analysis of compounds	Limited structural information, solvent interference, sensitive to temperature and pressure, can only analyze liquid samples, lower selectivity and sensitivity	Drug identification, quality control in a chemical research, food and beverage analysis
4	XRD	Fast, non-destructive, cost-effective, versatile, powerful, minimal sample preparation	Time consuming especially when analyzing large samples, limited to reference patterns, requires only a homogenous sample	Geological analysis, nuclear science, Nanotechnology, Cement industry
5	XRF	Speedy, accurate, non-destructive, portable, easy in ease	Interferences, background noise, matrix effects, limited depth of analysis	Medical applications, food inspection, archeological applications in stone analysis
6	NMR	Reproducible, quantitative, detailed, non-destructive, dynamic	Low magnetic moments for some elements, limited isotope abundance, equipment interference, magnetic field drift effect	Biomolecular analysis, Environmental analysis, pharmaceutical industry in drug analysis,
7	TEM	Detailed imaging, 3D mapping, high magnification and resolution.	Expensive in purchasing and maintenance, specialized experts required, inability to visualize living specimens	Nanotechnology, drug discovery, forensic analysis, semiconductor research
8	SEM-EDX	Rapid and high-resolution imaging, non-destructive, fast and cheap	Lack of specificity, limited to solid samples, poor accuracy to quantitative measurements of light elements	Materials science, forensics, pharmaceutical sciences, nanotechnology

REFERENCES

- Wang, J., Núñez, J., Maxwell, E. and Algar, W. (2015). Build your own photometer: A guided-inquiry experiment to introduce analytical instrumentation. *Journal of Chemical Education*, 4: 1-6.
- Bisen, P. and Sharma, A. (2013). Introduction to instrumentation in life sciences. *Taylor & Francis*, 1: 1-36. ISBN-13: 978-1-4665-1241-2.
- Vickerman, J. and Gilmore, I. (2009). Surface Analysis – The Principal techniques. *John Wiley & Sons, Ltd.* ISBN: 978-0-470-01763-0.
- Robinson, J.W., Frame, E. and Frame II, G. (2014). Undergraduate instrumental analysis. *Taylor & Francis*, ISBN-13: 978-1-4822-3374-2.
- Rayson, G.D. (2004). A unifying description of modern analytical instrumentation within a course on instrumental methods of analysis. *Journal of Chemical Education*, 81 (12): 1767-1771.
- Warner, D., Brown, E. and Shadle, S. (2016). Laboratory

- instrumentation: An exploration of the impact of instrumentation on student learning. *Journal of Chemical Education*, 93: 1223-1231.
- Kovarik, M., Clapis, J. and Romano-Pringle, A. (2020). Review of student-built spectroscopy instrumentation projects. *Journal of Chemical Education*, 97: 2185-2195.
- Shah, N., Arain, M. and Soylak, M. (2020). Historical background: milestones in the field of development of analytical instrumentation. New generation green solvents for separation and preconcentration. *Elsevier*, 2: 45-73.
- Farmer, D.K. and Jimenez, J.L. (2010). Real-time atmospheric chemistry field instrumentation. *Analytical Chemistry*, 82: 7879-7884.
- Liu, X., Deng, R., Zhang, Y., Wang, Y., Chang, H., Huang, L. and Liu, X. (2014). Probing the nature of up conversion nanocrystals: instrumentation matters. *Chemical Society Reviews*, 4: 1-30.
- Kumar, N., Sarma, A. and Rawal, R. (2014). Chemometrics tools used in analytical chemistry: An overview. *Talanta*, 123: 186-199.
- Tinega, J., Omosa, I. and Letema, S. (2023). Potential removal of chromium (VI) ions by macadamia nutshell from steel industry wastewater. *Environmental Engineering and Management Journal*, 22 (3): 411-427.
- Hummadi, K.K., Luo, S. and He, S. (2022). Adsorption of methylene blue dye from the aqueous solution via bio-adsorption in the inverse fluidized-bed adsorption column using the torrefied rice husk. *Chemosphere*, 287: 1-12.
- Rosanti, A.D., Kusumawati, Y., Hidayat, F., Fadlan, A., Wardani, A. and Agusyanti, H.A. (2022). Adsorption of methylene blue and methyl orange from aqueous solution using orange peel and CTAB-modified orange peel. *Journal of the Turkish Chemical Society*, 9 (1): 237-246.
- Deng, X., Xiong, F., Li, X., Xiang, B., Li, Z., Wu, X., Guo, C., Li, X., Li, Y., Li, G., Xiong, W. and Zeng, Z. (2018). Application of atomic force microscopy in cancer research. *Journal of Nanobiotechnology*, 16: 102-116.
- Njeri, J.N., Nthiga, E.W. and Muthakia, G. (2023). The efficacy of coffee husks biochar in the adsorption of methyl red from textile dyeing wastewater. *International Journal of Scientific Research in Chemical Sciences*, 10 (4):1-9.
- Oke, N and Mohan, S. (2022). Development of nanoporous textile sludge based adsorbent for the dye removal from industrial textile effluent. *Journal of Hazardous Materials*, 422: 1-12.
- Jaime, L., Dalia, I., Reyna, G. and Ma, A. (2018). Study of a fixed-bed column in the adsorption of an azo dye from an aqueous medium using a chitosan–glutaraldehyde biosorbent. *Adsorption Science & Technology*, 36 (1): 215-232.
- Ndung'u S.N., Wanjau R.N., Nthiga E.W., Ndiritu J. and Mbugua G.W. (2020). Complexation equilibrium studies of Cu²⁺, Cd²⁺ and Pb²⁺ ions onto ethylenediamine quaternised *Artocarpus heterophyllus* L. seeds from aqueous solution. *IOSR Journal of Applied Chemistry*, 13 (12): 1-12.
- Karthikeyan, S., Amudha, C., Tamilselvi, C. and Gopal, K. (2016). Removal of textile dye using carbon nanotubes as an adsorbent in fixed bed column. *Journal of Environmental Nanotechnology*, 5 (2): 17-24.
- Khan, N., Ryu, K.Y., Choi, J.Y., Nho, E.Y., Habte, G. and Choi, H. (2015). Determination of toxic heavy metals and speciation of arsenic in seaweeds from South Korea. *Food Chemistry* 169: 464-470.
- Ndung'u, S.N., Nthiga, E.W. and Wanjau, R.N. (2023). Characterization of amorphous silicon nitride prepared from sand and coffee husk wastes by carbothermal reduction-nitridation. *African Journal of Pure and Applied Sciences*, 4 (3): 24-34.
- Akash, M. and Rehman, K. (2020). Essentials of pharmaceutical analysis: Thermo Gravimetric Analysis. *Springer*, 19: 215-222.
- Potts, P.J., Ellis A.T., Kregsamer P., Marshall J., Streli C., West M. and Wobrauschek P. (2004) Atomic spectrometry update, X-ray spectrometry. *Journal of Analytical Atomic Spectrometry*, 19: 1397-1419.
- Ivanenko, N., Solovyev, N., Ivanenko, A. and Ganeev, A. (2012). Application of Zeeman Graphite Furnace Atomic Absorption Spectrometry with high-frequency modulation polarization for the direct determination of aluminum, beryllium, cadmium, chromium, mercury, manganese, nickel, lead, and thallium in human blood. *Archives of Environmental Contamination and Toxicology*, 1-10.
- Farrukh, M.A. (2012). Atomic Absorption Spectroscopy. *Intechopen*, 1: 1-270. ISBN 978-953-307-817-5.
- Wilberforce J.O. (2016). Review of principles and application of AAS, PIXE and XRF and their usefulness in environmental analysis of heavy metals. *IOSR Journal of Applied Chemistry*, 9 (6): 15-17.
- M Series Atomic Absorption Spectrometers (2005). Operators Manual, in *www.thermo.com*, Issue 2.
- Paul, B.N., Chanda, S., Das, S., Singh, P., Pandey, B. and Giri, S. (2014). Mineral assay in Atomic Absorption Spectroscopy. *The Beats of Natural Sciences*, 1 (4): 1-17.
- Paul, V., Pandey, R., Ramesh, K. and Pal, M (2017). Atomic Absorption Spectroscopy (AAS) for Elemental Analysis of Plant Samples. In manual of ICAR Sponsored Training Programme for Technical Staff of

- ICAR Institutes on "Physiological Techniques to Analyze the Impact of Climate Change on Crop Plants". Division of Plant Physiology, ICAR-Indian Agricultural Research Institute (IARI), New Delhi, India.
- Sudunagunta, D., Venkatesh. D.N. and Meyyanathan, S.N. (2012). Atomic Absorption Spectroscopy: A special emphasis on pharmaceutical and other applications. *Journal of Pharmacy Research*, 5 (3): 1614-1619.
- Yeung, V., Miller, D. and Rutzke, M. (2017). Atomic absorption spectroscopy, atomic emission spectroscopy, and inductively coupled plasma-mass spectrometry. *Springer*, 9: 129-150.
- Sharma, B. and Tyagi, S. (2013). Simplification of metal ion analysis in fresh water samples by Atomic Absorption Spectroscopy for laboratory students. *Journal of Laboratory Chemical Education*, 1 (3): 54-58.
- Haas, J. and Mizaikoff, B. (2016). Advances in mid-infrared spectroscopy for chemical analysis. *Annual Review of Analytical Chemistry*, 9: 45-68.
- Amir, R., Anjum, F., Khan, M., Khan, M., Pasha, I. and Nadeem, M. (2013). Application of Fourier transform infrared (FTIR) spectroscopy for the identification of wheat varieties. *Journal of Food Science and Technology*, 50 (5): 1018-1023.
- Kurniawati, D., Bahrizal, Sari, T., Adella, F. and Sy, S. (2021). Effect of contact time adsorption of Rhodamine B, Methyl Orange and Methylene Blue colours on langsat shell with batch methods. *Journal of Physics: Conference Series*, 1788: 1-6.
- Cocchi, M., Foca, G., Lucisano, M., Marchetti, A., Paeon, M.A., Tassi, L. and Ulrici, A. (2004) Classification of cereal flours by chemometric analysis of MIR spectra. *Journal of Agricultural and Food Chemistry*, 52: 1062-1067.
- Munajad, A., Subroto, C. and Suwarno, S. (2018). Fourier Transform Infrared (FTIR) spectroscopy analysis of transformer paper in mineral oil-paper composite insulation under accelerated thermal aging. *Energies*, 11 (2): 364-375.
- Al-Ghouti, M. and Al-Absi, R. (2020). Mechanistic understanding of the adsorption and thermodynamic aspects of cationic methylene blue dye onto cellulosic olive stones biomass from wastewater. *Scientific Reports*, 10: 1-18.
- Sivakumar, S., Ravisankar, R., Raghu, Y., Chandrasekaran, A. and Chandramohan, J. (2012). FTIR spectroscopic studies on coastal sediment samples from Cuddalore District, Tamilnadu, India. *Indian Journal of Advances in Chemical Science*, 1: 40-46.
- Chen, Y., Furmann, A., Mastalerz, M. and Schimmelmann, A. (2014). Quantitative analysis of shales by KBr-FTIR and micro-FTIR. *Fuel*, 116: 538-549.
- Cui, H., Abu-Siada, A., Li, S. and Islam, S. (2017). Correlation between dissolved gases and oil spectral response. *1st international conference on electrical materials and power equipment*. IEEE. Pp. 28-32.
- Țucureanu, V., Matei, A. and Avram, A. M. (2016). FTIR spectroscopy for carbon family study. *Critical Reviews in Analytical Chemistry*, 46 (6): 502-520.
- Perez-Guaita, D., Andrew, D., Heraud, P., Beeson, J., Anderson, D., Richards, J., and Wood, B. (2016). High resolution FTIR imaging provides automated discrimination and detection of single malaria parasite infected erythrocytes on glass. *Faraday discussions*, 187: 341-352.
- Faghihzadeh, F., Anaya, N., Schiffman, L. and Oyanedel-Craver, V. (2016). Fourier Transform Infrared Spectroscopy to assess molecular-level changes in microorganisms exposed to nanoparticles. *Nanotechnology for Environmental Engineering*, 1 (1): 1-16.
- Gonciarz, L., Urbaniak, M., Kaca, W. and Chmiela, M. (2020). Use of Fourier-Transform Infrared spectroscopy (FT-IR) for monitoring experimental *Helicobacter pylori* infection and related inflammatory response in guinea pig model. *International Journal of Molecular Sciences*, 22 (1): 281-297.
- Santhoshkumar, J., Kumar, S. and Rajeshkumar, S. (2017). Synthesis of zinc oxide nanoparticles using plant leaf extract against urinary tract infection pathogen. *Resource-Efficient Technologies*, 3: 459-465.
- Simonescu, C. (2012). Application of FTIR spectroscopy in environmental studies. *InTech*, 2: 49-84.
- Valand, R., Tanna, S., Lawson, G. and Bengtström, L. (2019). A review of Fourier Transform Infrared (FTIR) spectroscopy used in food adulteration and authenticity investigations. *Food Additives & Contaminants: Part A*, 1-21.
- Chakraborty, S., Sharmin, S., Rony, S. R., Ahmad, S. and Sohrab, M. (2018). Stability-indicating UV/Vis spectrophotometric method for diazepam, development and validation. *Indian Journal of Pharmaceutical Sciences*, 80 (2): 366-373.
- Piccollo, M., Aceto, M. and Vitorino, T. (2018). UV-Vis spectroscopy. *Physical Sciences Reviews*, 1-14.
- Verma, G. and Mishra, M. (2018). Development and optimization of UV-Vis spectroscopy-A review. *World Journal of Pharmaceutical Research*, 7 (11): 1170-1180.
- Singh, K. (2020). Spectrophotometer (UV-Visible). *Compendium of Biomedical Instrumentation*, 343: 1815-1824.
- Haddi, R., El Kharraz, A. and Kerroumi, M.I. (2024). Green synthesis of zinc oxide nanoparticles using *Pistacia lentiscus* L. leaf extract and evaluating their antioxidant and antibacterial properties. *Nano*

- Biomedicine and Engineering*, 16 (2): 232-247.
- Guo, Y., Liu, C., Ye, R. and Duan, Q. (2020). Advances on water quality detection by UV-Vis spectroscopy. *Applied Sciences*, 10: 6874-6891.
- Abu Bakar, N., Cui, H., Abu-Siada, A. and Li, S. (2016). A Review of Spectroscopy Technology Implementation in Transformer Condition Monitoring. *Proceedings of the International Conference on Condition Monitoring and Diagnosis*, (CMD2016), 25-28 Sept 2016, Xi'an, China.
- Rocha, F., Gomes, A., Lunardi, C., Kaliaguine, S. and Patience, G.S. (2018). Experimental methods in chemical engineering: Ultraviolet visible spectroscopy|UV-Vis. *The Canadian Journal of Chemical Engineering*, 96 (12): 2512-2517.
- Shinde, G., Godage, R.K., Jadhav, R.S., Manoj, B. and Aniket, B. (2020). A Review on Advances in UV Spectroscopy. *Research Journal of Science and Technology*, 12 (1): 47-51.
- Khan, H., Yerramilli, A., D'Oliveira, A., Alford, T., Boffito, D. and Patience, G. S. (2020). Experimental methods in chemical engineering: X-ray diffraction spectroscopy-XRD. *Canadian Journal of Chemical Engineering*, 98 (6): 1255-1266.
- Bishnoi, A., Kumar, S. and Joshi, N. (2017). Wide-angle X-ray Diffraction (WXRD): Technique for characterization of nanomaterials and polymer nanocomposites. In *microscopy methods in nanomaterials characterization*. Elsevier, 9: 313-337.
- Titus, D., Samuel, E. and Roopan, S. (2019). Nanoparticle characterization techniques. In *Green Synthesis, Characterization and Applications of Nanoparticles*. Elsevier, 12: 303-319.
- Fakhirah, D., Maghfira, T., Hutama, A., Septama, A., Maryani, F. and Krismastuti, F. (2024). Synthesis, characterization, and antibacterial activity of plant-derived zinc oxide nanostructure using *Lavandula angustifolia* and *Phyllanthus niruri* extracts. *Indonesian Journal of Chemistry*, 24 (3): 865-875.
- Melkamu, W. and Feleke, E. (2022). Green synthesis of copper oxide nanoparticles using leaf extract of *Justicia Schimperiana* and their antibacterial activity. *Research Square*, 1-24.
- El-Azazy, M., El-Shafie, A., Al-Mulla, R., Hassan, S. and Nimir, H. (2023). Enhanced adsorptive removal of rifampicin and tigecycline from single system using nano-ceria decorated biochar of mango seed kernel. *Heliyon*, 9: 1-21.
- Pandey, A., Dalal, S., Dutta, S. and Dixit, A. (2021). Structural characterization of polycrystalline thin films by X-ray diffraction techniques. *Journal of Materials Science: Materials in Electronics*, 32:1341-1368.
- Ermrich, M. and Opper, D. (2011). XRD for the analyst. *PANalytical GmbH*. ISBN: 978-90-809086-0-4
- Bunaciu, A., Udristoiu, E. and Aboul-Enein, H. (2015). X-Ray Diffraction: Instrumentation and applications. *Critical Reviews in Analytical Chemistry*, 45 (4): 289-299.
- Epp, J. (2016). X-ray diffraction (XRD) techniques for materials characterization. *Materials Characterization Using Nondestructive Evaluation (NDE) Methods*. Elsevier, 4: 81-124.
- Kim, S., Lee, C. and Kafle, K. (2013). Characterization of crystalline cellulose in biomass: Basic principles, applications, and limitations of XRD, NMR, IR, Raman, and SFG. *Korean Journal of Chemical Engineering*, 1-15.
- Cornelius T.W. and Thomas, O. (2018). Progress of in situ synchrotron X-ray diffraction studies on the mechanical behavior of materials at small scales. *Progress in Materials Science*, 94: 384-434.
- George, F. and José, S. (2021). Back-to-Basics tutorial: X-ray diffraction of thin films. *Journal of Electroceramics*, 1-33.
- Das, R., Ali, M. and Hamid, S. (2014). Current applications of X-Ray powder diffraction- A Review. *Review on Advanced Materials Science*, 38 (2): 95-109.
- Plaipichit, S., Wicharn, S., Puttharugsa, C., Wanakamol, P. and Buranasiri, P. (2018). Virtual X-ray diffractometer using acoustic wave for material science education. *IOP Conference Series: Journal of Physics*, 1144 (1): 1-5.
- Muniz, F., Miranda, M., dos Santos, C. and Sasaki, J. (2016). The Scherrer equation and the dynamical theory of X-ray diffraction. *Acta Crystallographica Section A*, 72: 385-390.
- Chen, C., Sun, K., Huang, C., Yang, M., Fan, M., Wang, A., Zhang, G., Li, B., Jiang, J., Xu, W. and Liu, J. (2023). Investigation on the mechanism of structural reconstruction of biochars derived from lignin and cellulose during graphitization under high temperature. *Biochar*, 5 (1): 51-64.
- Grieken, R.V. and Margui', E. (2013). X-ray fluorescence spectrometry and related techniques: An introduction, momentum Press, New York.
- Potter, N. and Brand, N. (2019). Application of micro-XRF to characterize diamond drill-core from lithium-caesium-tantalum pegmatites. *ASEG Extended Abstracts*, 1: 1-4.
- Oyedotun, T. (2018). X-ray fluorescence (XRF) in the investigation of the composition of earth materials: a review and an overview. *Geology, Ecology, and Landscapes*, 2 (2): 148-154.
- Tsuji, K., Matsuno, T., Takimoto, Y., Yamanashi, M., Kometani, N., Sasaki, Y., Hasegawa, T., Kato, S., Yamada, T., Shoji, T. and Kawahara, N. (2015). New developments of X-ray fluorescence imaging techniques

- in laboratory. *Spectrochimica Acta Part B*, 113: 43-53.
- Marguí, E. and Grieken, R. (2013). State-of-the-art X-ray Fluorescence instrumentation for chemical analysis. *XRF Sulphur Analysis Spotlight*, 16: 1-3.
- Gupta, A.K. (2014). Total Reflection X-Ray Fluorescence spectroscopy –working principles. *International Journal of Core Engineering & Management*, 1 (5): 36-55.
- Montanha, G., Rodrigues, E., Marques, J., de Almeida, E., dos Reis, A. and de Carvalho, H. (2020). X-ray fluorescence spectroscopy (XRF) applied to plant science: challenges towards in vivo analysis of plants. *Metallomics*, 12: 183-192.
- Marguí, E., Zawisza, B. and Sitko, R. (2014). Trace and ultratrace analysis of liquid samples by X-Ray fluorescence spectrometry. *Trends in Analytical Chemistry*, 53: 73-83.
- Shackley, M. (2011). An introduction to X-Ray Fluorescence (XRF) analysis in geoarchaeology. *Springer*, 2: 7-44.
- Bothwell, J. and Griffin, J. (2011). An introduction to biological nuclear magnetic resonance spectroscopy. *Biological Reviews*, 86: 493-510.
- Claridge, T.D. (2016). High-Resolution NMR techniques in organic chemistry. *Elsevier*, 27: 1-10.
- Yu, H.-Y., Myoung, S. and Ahn, S. (2021). Recent Applications of Benchtop Nuclear Magnetic Resonance Spectroscopy. *Magnetochemistry*, 7: 121-146.
- Muller, C., Kong, X., Cai, J., Melentijevic', K., Stacey, A., Markham, M., Twitchen, D., Isoya, J., Pezzagna, S., Meijer, J., Du, J.F., Plenio, M.B., Naydenov, B., McGuinness, L.P. and Jelezko, F. (2014). Nuclear magnetic resonance spectroscopy with single spin sensitivity. *Nature Communications*, 5: 4703-4708.
- Simpson, A., Simpson, M. and Soong, R. (2018). Environmental Nuclear Magnetic Resonance spectroscopy: An overview and a primer. *Analytical Chemistry*, 90: 628-639.
- Mishra, R.K., Cherusseri, C., Bishnoi, A. and Thomas, S. (2017). Nuclear Magnetic Resonance Spectroscopy. Spectroscopic methods for nanomaterials characterization. *Elsevier*, 13: 369-415.
- Sugiki, T., Kobayashi, N. and Fujiwara, T. (2017). Modern technologies of solution Nuclear Magnetic Resonance Spectroscopy for three-dimensional structure determination of proteins open avenues for life scientists. *Computational and Structural Biotechnology Journal*, 15: 328-339.
- Simpson, A., Simpson, M. and Soong, R. (2012). Nuclear Magnetic Resonance spectroscopy and its key role in environmental research. *Environmental Science & Technology*, 46: 11488-11496.
- Espina, R., Yu, L., Wang, J., Tong, Z., Vashishtha, S., Talaat, R., Scatina, J. and Mutlib, A. (2009). Nuclear Magnetic Resonance spectroscopy as a quantitative tool to determine the concentrations of biologically produced metabolites: Implications in metabolites in safety testing. *Chemical Research in Toxicology*, 22 (2): 299-310.
- Kong, X., Stark, A., Du, J., McGuinness, L. and Jelezko, F. (2015). Towards chemical structure resolution with nanoscale Nuclear Magnetic Resonance spectroscopy. *Physical Review Applied*, 4: 1-6.
- Zia, K., Siddiqui, T., Ali, S., Farooq, I., Zafar, M. and Khurshid, Z. (2019). Nuclear Magnetic Resonance spectroscopy for medical and dental applications: A Comprehensive review. *European Journal of Dentistry*, 13 (1): 124-128.
- Keeler, J. (2011). Understanding NMR spectroscopy. Chichester: John Wiley & Sons.
- Kaech, A. (2013). An introduction to Electron Microscopy instrumentation, imaging and preparation. Center for Microscopy and Image Analysis, University of Zurich. 1-28.
- Franken, L., Boekema, E. and Stuart, M. (2017). Transmission Electron Microscopy as a tool for the characterization of soft materials: application and interpretation. *Advanced Science News*, 600476: 1-9.
- Geetanjali, Tamta, A., Chandra, B., Kandpal, N.D. and Joshi, R. (2024). Green route synthesis of manganese oxide nanoparticles by using methanolic extract of *Sapindus mukorossi* (reetha). *Journal of Water and Environmental Nanotechnology*, 9 (2): 211-222.
- Goodhew, P. (2011). General introduction to Transmission Electron Microscopy (TEM). Aberration-Corrected Analytical Transmission Electron Microscopy. *John Wiley & Sons*, 1: 1-20.
- Inkson, B.J (2016). Scanning Electron Microscopy (SEM) and Transmission Electron Microscopy (TEM) for materials characterization. Materials characterization using nondestructive evaluation (NDE) methods. *Elsevier*, 2:18-43.
- Reimer, L. and Kohl, H. (2008). Transmission Electron Microscopy: Physics of Image Formation, fifth ed. Springer. ISBN: 13: 978-0387400938.
- Nie, B., Liu, X., Yang, L., Meng, J. and Li, X. (2015). Pore structure characterization of different rank coals using gas adsorption and scanning electron microscopy. *Fuel*, 158: 908-917.
- Han, W., Jiao, H. and Fox, D. (2018). Scanning Electron Microscopy. In: *Springer Tracts in Modern Physics*, 272: 35-68.
- Sharma, R., Patel, K.S., Lata, L. and Milosh, H. (2016). Characterization of Urban Soil with SEM-EDX. *American Journal of Analytical Chemistry*, 7: 724-735.
- Cardell, C. and Guerra, I. (2015). An overview of emerging hyphenated SEM-EDX and Raman spectroscopy

- systems: Applications in life, environmental and materials sciences. *Trends in Analytical Chemistry*, 1-39.
- El-Zahhar, A., Idris, A., Fawy, K. and Arshad, M. (2019). SEM, SEM-EDX, μ -ATR-FTIR and XRD for urban street dust characterisation. *International Journal of Environmental Analytical Chemistry*, 1-20.
- Haddi, R., El Kharraz, A.M. and Kerroumi, M.I. (2024). Green synthesis of zinc oxide nanoparticles using pistacia lentiscus L. leaf extract and evaluating their antioxydant and antibacterial properties. *Nano Biomedicine and Engineering*, 16 (2): 232-247.
- Abd Motalib, M., Rahman, M.A., Othman M., Ismail, A. and Jaafar, J. (2017). Scanning Electron Microscopy (SEM) and Energy Dispersive X-Ray (EDX) spectroscopy. In membrane characterization. *Elsevier*, 9: 161-179.